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Appl. No. 10/053,262 Amendment dated July 26, 2004 Reply to Office Action of February 26, 2004

REMARKS/ARGUMENTS

- 1. Claims 1-3 are pending.
- 2. Claims 1-2 stand rejected, under 35 U.S.C. § 103(a), for obviousness over Cheung et al. (hereinafter "Cheung"), in view of Schuurs et al. (hereinafter "Schuurs"), and further in view of Metcalfe et al. (hereinafter "Metcalfe").

Applicants submit herewith Exhibit A, which details that the Cheung reference (Analytical Chemistry 282: 24-28, 2000) was published in the June 15, 2000 edition of the journal. Therefore, the Cheung reference does not qualify as prior art, under 35 U.S.C. § 102(b), over the present application, which claims priority from U.S. Provisional Patent Appl. No. 60/265,135, filed January 30, 2001.

Furthermore, Applicants submit herewith a 37 C.F.R. 1.131 Declaration of Janice A. Brown, co-inventor on the present application, which recounts that the claimed invention of claims 1 and 2 was conceived and reduced to practice prior to June, 2000.

Accordingly, Applicants respectfully request reconsideration of the Office Action mailed February 26, 2004.

3. Claim 3 stands rejected, under 35 U.S.C. § 103(a), for obviousness over Cheung, in view of Schuurs, in further view of Metcalfe, and further in view of Craig et al. (hereinafter "Craig").

Applicants submit herewith a 37 C.F.R. 1.131 Declaration of Janice A. Brown, coinventor on the present application, which describes that the claimed invention of claim 3 was conceived and reduced to practice prior to June, 2000.

Accordingly, Applicants respectfully request reconsideration of the Office Action mailed February 26, 2004.

4. In view of the Remarks/Arguments hereinabove, and the Declaration submitted herewith, Applicants believe that the Application is in condition for immediate allowance. Therefore, reconsideration of the Office Action mailed February 26, 2004, and a timely Notice of Allowance is respectfully requested.

Appl. No. 10/053,262 Amendment dated July 26, 2004 Reply to Office Action of February 26, 2004

Data: 7.1 26th 2004

Deborah A. Martin Attorney for Applicant Reg. No. 44,222

Respectfully submitted,

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Patent Department, MS 8260-1611
Eastern Point Road
Groton, Connecticut 06340
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Exhibit A

	EXIIIDIL A	
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	rages 1-9 Heike Borcherding, Steven Leikefeld, Christa Frey, Stephan Diekmann and Peter Steinrücke Abstract Abstract + References PDF (99 K)	Diekmann and Peter Steinrücke
2.	Determination of the Binding Parameters of Drug to Protein by Equilibrium Dialysis/Piezoelectric Quartz Crystal	ein by Equilibrium Dialysis/Piezoelectric Ouartz Crystal
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<u> </u>	High-Resolution Magic Angle Spinning ¹ H NMR Spectroscopy of Intact Liver and Kidney: Optimization of Sample Preparation Procedures and Biochemical Stability of Tissue during Spectral Acquisition • ARTICLE	copy of Intact Liver and Kidney: Optimization of Sample ne during Spectral Acquisition • ARTICLE
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6. [7]	Catalytic Chromatography • ARTICLE Pages 39-45 Luis A. Jurado, James T. Drummond and Harry W. Jarrett Abstract Abstract + References PDF (124 K)
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12.	Determination of Ascorbic Acid and Dehydroascorbic Acid in Biological Samples by High-Performance Liquid Chromatography Using Subtraction Methods: Reliable Reduction with Tris[2-carboxyethyl]phosphine Hydrochloride • ARTICLE Pages 89-93 Jens Lykkesfeldt Abstract Abstract + References PDF (67 K)
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Purpose: Repeat the compound (see be bu) dose responses using the biotinylated NAD PARP ELISA assay format

Procedure

Biotinglated NAD

PARP (partially parified) 5 ug/l
5.3 mg/l protein in stock
clilute stock 1:1000 to make a 5.3 ug/l stock
For I plt make 13.1 huffer (50mm to 1101)

13 ulo PARP+ 12987 I buffer (50mm Tris HCl pH 8.0, 20mm 2nch

Coat Plates overnight at 4°C

Wash 3x w T-PBS

Add the following (wote for the next step keep all reagents on ice)

Buffer (50mm Tris HCl pH 80, 20mm 2nCl2 4mm mgCl2)
For 1 pH make 15mls buffer (50ml 2nCl2 4mm mgCl2)
Cold NAD 100mm (FC)
W. 34mg L = 100mm NAD
Low 34mg = 100mm NAD
1000 1 15 l buffer = 100mm NAD

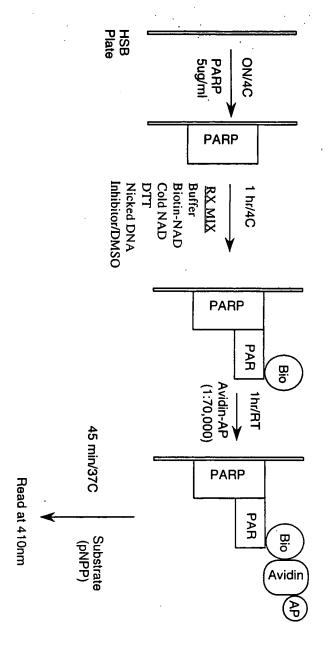
DIT Imm (FC) 1524mg/L=1mm DTT 1524mg = 12 = 2286mg DTT/15-1 buffer=1mmDTT

Biotinulated NAD (25mm RC)
Make 14.5 I so add
1.450 I Biotinulated NAD to 13.050 I of buffer
Containing cold NAD (100mm) and DIT (1mm)
Note: Adding biotinulated NAD to the above
told NAD and + DTI buffer will cause a
1070 reduction in buffer cone so now have
15mm Tris Hel, 18mm 2ncl2, 3.6mm macl2
90mm cold NAD and 900mm DTT. This was the
Same way buffer establishing these assay

Janice Brown

Hereful Mae Judien

PARP Biotinylated NAD Assay



Jose O Mayore

Jane Brown



I hereby certify that this correspondence is being deposited with the United States Postal Service as first-class mail in an envelope addressed to: Hon. Commissioner for Patents, PO Box 1450, Alexandria, VA 22313-1450 on this 26th day of July, 2004.

By

(Signature of person mailing)

Janice M. Denison

(Typed or printed name of person)

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Appl. No.: 10/053,262

In Re Application of Janice A. Brown

et al.

Filed: January 18, 2002 Group Art Unit: 1641

Examiner: Davis, Deborah A. Docket No.: PC11044ADAM

Customer No.: 28523

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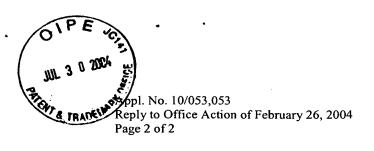
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37 CFR § 1.131 DECLARATION OF JANICE A. BROWN

Sir:

- 1. I, Janice A. Brown, am the first named inventor of the above-captioned application.
- 2. I, together with co-inventor Ravi B. Marala, conceived of, and reduced to practice, the invention claimed in Claims 1-3 of the pending application, before June, 2000.
- 3. Exhibit B, attached herewith, is a copy of a laboratory notebook (2 pages total) that I prepared, signed, and had witnessed, before June, 2000. Exhibit B is redacted only as to the dates of the laboratory notebook disclosure and witnessing, dates which all occur before June, 2000.
- 4. Exhibit A describes a method to assay poly(ADP-ribose) polymerase (PARP) activity which includes the steps of immobilizing PARP on a multiwell HSB plate, contacting the immobilized PARP with biotinylated nicotinamide-adenine-dinucleotide (NAD), at 4°C, and under conditions that allow PARP auto-ribosylation, contacting the auto-ribosylated PARP with



an avidin-conjugated alkaline phosphatase detectable marker, thereby forming a complex between the auto-ribosylated PARP and the detectable marker, and measuring the amount of detectable marker complexed to auto-ribosylated PARP using a p-nitrophenyl phosphate (pNPP) substrate of alkaline phosphatase as an indication of the amount of PARP activity in the sample, wherein none of the above-listed reagents are radioactive.

5. I further declare that all statements made herein of my own knowledge are true and that all statements made on belief are believed true; and further, that these statements were made with knowledge that willful false statements and the like so made are punishable by fine or imprisonment or both, under Section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the above-captioned application or any patent issuing therefrom.

Date

Janice A. Brown